

Review

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Astaxanthin in microalgae: pathways, functions and biotechnological implications

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Major progress has been made in the past decade towards understanding of the biosynthesis of red carotenoid astaxanthin and its roles in stress response while exploiting microalgae-based astaxanthin as a potent antioxidant for human health and as a coloring agent for aquaculture applications. In this review, astaxanthin-producing green microalgae are briefly summarized with *Haematococcus pluvialis* and *Chlorella zofingiensis* recognized to be the most popular astaxanthin-producers. Two distinct pathways for astaxanthin synthesis along with associated cellular, physiological, and biochemical changes are elucidated using *H. pluvialis* and *C. zofingiensis* as the model systems. Interactions between astaxanthin biosynthesis and photosynthesis, fatty acid biosynthesis and enzymatic defense systems are described in the context of multiple lines of defense mechanisms working in concert against photooxidative stress. Major pros and cons of mass cultivation of *H. pluvialis* and *C. zofingiensis* in phototrophic, heterotrophic, and mixotrophic culture modes are analyzed. Recent progress in genetic engineering of plants and microalgae for astaxanthin production is presented. Future advancement in microalgal astaxanthin research will depend largely on genome sequencing of *H. pluvialis* and *C. zofingiensis* and genetic toolbox development. Continuous effort along the heterotrophic-phototrophic culture mode could lead to major expansion of the microalgal astaxanthin industry.

Key Words: astaxanthin biosynthesis; *Chlorella zofingiensis*; genetic engineering; *Haematococcus pluvialis*; mass culture; photooxidative stress

INTRODUCTION

Astaxanthin (3,3'-dihydroxy- β,β -carotene-4,4'-dione) is a ketocarotenoid synthesized by limited numbers of microalgae, plants, bacteria, and fungi. In microalgae, unlike primary carotenoids (e.g., β -carotene, zeaxanthin, and lutein) which constitute structural and functional components of the photosynthetic apparatus, astaxanthin is a secondary carotenoid accumulating in cytosolic lipid bodies (LBs) only under environmental stress or adverse culture conditions, such as high light, high salinity,

and nutrient deprivation (Johnson and An 1991, Johnson and Schroeder 1996).

Astaxanthin has thirteen conjugated double bonds arranged as alternating single-double bonds. This configuration confers astaxanthin strong antioxidant properties thereby scavenging reactive oxygen species (ROS) and neutralizing free radicals (Miki 1991, Vershinin 1999). Because of this, astaxanthin has been used as a nutraceutical and a pharmaceutical, for example, to fight against

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free-radical associated diseases like oral, colon and liver cancers, cardiovascular diseases, and degenerative eye diseases (Lorenz and Cysewski 2000, Guerin et al. 2003). Astaxanthin is also a common coloring agent in aquaculture to impart red pigmentation in animal bodies such as salmon and rainbow trout (Lorenz and Cysewski 2000, Guerin et al. 2003).

The current world astaxanthin market is dominated by synthetic astaxanthin. The price for synthetic astaxanthin is above \$2,000 kg⁻¹, and the total market value of astaxanthin is over \$240 M per year (Misawa 2009, Li et al. 2011). Synthetic astaxanthin typically contains a mixture of 3S, 3'S; 3R, 3'S; and 3R, 3'R isoforms with a ratio of 1 : 2 : 1. In contrast, astaxanthin from microalgae is predominantly the 3S, 3'S isomer. The 3S, 3'S astaxanthin isomer is a preferable form as a feed additive in aquaculture because it imparts a higher extent of pigmentation in rainbow trout than the synthetic astaxanthin (Barbosa et al. 1999). There are growing concerns about the safety of using synthetic astaxanthin for aquaculture or direct human consumption and therefore natural astaxanthin represents a preferred and premium choice for human consumption (Li et al. 2011).

Astaxanthin can be produced in various amounts by a number of microalgae including *Botryococcus braunii* (up to 0.01% astaxanthin by dry weight (dwt)) (Grung et al. 1994), *Chlamydocapsa* spp. (0.04% astaxanthin by dwt) (Leya et al. 2009), *Chlamydomonas nivalis* (Bidigare et al. 1993, Remias et al. 2005), *Chlorella zofingiensis* (0.7% astaxanthin by dwt) (Bar et al. 1995, Orosa et al. 2000), *Chlorococcum* sp. (0.7% astaxanthin by dwt) (Liu and Lee 2000, Ma and Chen 2001), *Chloromonas nivalis* (0.004% astaxanthin by dwt) (Leya et al. 2009, Remias et al. 2010), *Eremosphaera viridis* (Vechtel et al. 1992), *Haematococcus pluvialis* (4% astaxanthin by dwt) (Droop 1954, Lee and Ding 1994), *Neochloris wimmeri* (1.9% astaxanthin by dwt) (Orosa et al. 2000), *Protosiphon botryoides* (1.4% astaxanthin by dwt) (Orosa et al. 2000), *Scenedesmus* sp. (0.3% astaxanthin by dwt) (Orosa et al. 2000, Qin et al. 2008), *Scotiellopsis oocystiformis* (1.1% astaxanthin by dwt) (Orosa et al. 2000), and *Trachelomonas volvocina* (Green 1963). Of these species, the green microalga *H. pluvialis* is recognized as one of the most promising producer of astaxanthin in nature due to its exceptional ability to accumulate large amounts of astaxanthin under stress conditions (Boussiba 2000, Lemoine and Schoefs 2010). Recently, *C. zofingiensis* has attracted some interest as an alternative astaxanthin producer, due to its ability to grow rapidly (with a maximum specific growth rate of 1.03 d⁻¹ and biomass concentration of 53 g L⁻¹ when

grown on glucose) and adopt a photoautotrophic, mixotrophic, or heterotrophic culture mode (Orosa et al. 2000, Del Campo et al. 2004, Ip et al. 2004, Sun et al. 2008).

The biosynthesis of astaxanthin in microalgae is usually accompanied by the transformation of the algae from a green vegetative form into a red cyst (Droop 1954). The cellular and molecular responses of *H. pluvialis* and *C. zofingiensis* to stress are proposed to result, at least in part, from differential expression of carotenoid synthesis genes at the transcriptional level (Hershkovits et al. 1997, Li et al. 2008a, 2010). Recently, many genes responsible for carotenoids, particularly astaxanthin biosynthesis have been cloned and characterized in *H. pluvialis* and *C. zofingiensis*, which has provided the opportunities to study the pathways and regulation of carotenoid biosynthesis and understand the biological role of astaxanthin in stress response. This review summarizes the cell biology, physiological, and biochemical characteristics of *H. pluvialis* and *C. zofingiensis*, with emphasis on recent progress on the pathways for and the physiological role of astaxanthin synthesis under stress. The biotechnological implications of these studies are discussed and recent efforts on genetic engineering of algal genes in microalgae and higher plants for overproduction of astaxanthin are described.

HAEMATOCOCCUS PLUVIALIS

Cell biology and physiology

The life cycle of *H. pluvialis* consists of four types of distinguishable cells, including macrozooids (or zoospores), microzooids, palmella, and aplanospores (or haematocysts) (Elliot 1934). Macrozooids are spherical, ellipsoidal, or pear-shaped cells with two flagella and a cup-shaped chloroplast. Exhibited by rapid growth and cell division producing 2-8 daughter cells, macrozooids predominate in the early vegetative growing stage of a batch culture under favorable culture conditions. When macrozooids are subjected to unfavorable environmental or culture conditions, they develop into a non-motile 'palmella' form by losing their flagella while expanding the cell size. This process is defined as encystment. When the stress persists, 'palmella' will develop into the non-motile asexual aplanospores with thick and rigid cell walls. During the maturation of aplanospores, large amounts of astaxanthin accumulate, which brings a bright red color to the cells. It is worth noting that macrozooids of some *H. pluvialis* strains are capable of accumulating astaxan-

thin without forming aplanospores (Brinda et al. 2004, Hagen et al. 2004). If the environmental or culture conditions are back to normal, aplanospores will germinate to form flagellated zoospores to start a new vegetative growth cycle. In some cases, after exposure to extreme adverse conditions, e.g., freezing, desiccation, or nutrient starvation, followed by favorable culture conditions, gametogenesis may occur in aplanospores, giving rise to up to 64 gametes, known as microzooids. The microzooids appear as smaller cells (<10 µm) as compared to the zoospores (20-50 µm), and exhibit high-speed motility after their release from gametocysts (Triki et al. 1997).

Accumulation of astaxanthin in *H. pluvialis* cells is accompanied by profound ultrastructural changes. One of the most remarkable changes is the formation of a large number of cytosolic LBs where bulks of astaxanthin molecules are stored. Initially, LBs appear as tiny electron-dense spots scattered near the rough endoplasmic reticulum cisternae and eventually are coalesced to form large mature LBs (Lang 1968, Santos and Mesquita 1984).

Flagellated zoospores of *H. pluvialis* exhibit a voluminous extracellular matrix, resembling the typical Volvocan multilayered architecture with a tripartite crystalline layer between two separate layers of gelatinous matrices. The tripartite crystalline layer is connected to the cytoplasmic membrane by fine radiating strands, which is only observed in the genus of *Haematococcus*. Encystment of the flagellated zoospores into non-motile palmella cells is characterized by the formation of the primary cell wall within the extracellular matrix (Hagen et al. 2008). The primary cell wall can be stained with calcofluor white, indicative of the presence of β -1,4-glycosidic linkages. Further formation of red aplanospores is concurrent with the development of secondary cell wall, which contains 3% acetolysis-resistant materials suggested to be algaenan, an aliphatic biopolymer with great resistance to chemicals (Damiani et al. 2006, Hagen et al. 2008). Recently, the proteomes of *H. pluvialis* cell wall at different developmental stages were characterized, suggesting possible involvement of cellulose synthesis in the primary cell formation (Wang et al. 2004b).

The most noticeable response of *H. pluvialis* to adverse culture conditions is to synthesize and accumulate large amounts of astaxanthin in the form of mono- and di-esters. While the initial astaxanthin synthesis utilizes the existing β -carotene as a precursor, produce a bulk of astaxanthin esters will depend on *de novo* β -carotene synthesis (Schoefs et al. 2001).

Nearly 50% reduction in chlorophyll content occurred during the transformation of green vegetative cells into

red aplanospores (Boussiba et al. 1999, Han et al. 2012), and yet red aplanospores were photosynthetically active with only moderate declines in the maximal photosynthetic rate (P_{max}) and the maximum quantum yield of photosystem II (PSII), presumably attributable to the relatively stable photosynthetic unit size and the activated D1 repairing mechanism (Zlotnik et al. 1993, Wang et al. 2003, Qiu and Li 2006). Attenuated linear electron transport as indicated by significantly reduced cytochrome *b6f* content and the concomitant enhanced plastid terminal oxidase (PTOX)-mediated alternative electron transport were considered to prevent the photosynthetic electron transport chain from being over-reduced when *H. pluvialis* cells are under photooxidative stress (Tan et al. 1995, Han et al. 2012). On the other hand, the PSI activity was significantly enhanced during the transformation of green vegetative cells to red aplanospores (Hagen et al. 1993, Tan et al. 1995, Qiu and Li 2006).

CHLORELLA ZOFINGIENSIS

The unicellular microalgae *Chlorella* spp. have been widely used in basic biology research and various fields of biotechnology, such as animal feeds, health food and bioenergy production (Huss et al. 1999). *C. zofingiensis* (Chlorellaceae) has a rigid cell wall containing glucose and mannose, and is able to accumulate secondary carotenoids, whereas other species of *Chlorella*, such as *C. vulgaris*, have a glucosamine-containing cell wall and no capability of accumulating secondary carotenoids (Takeida 1991).

In response to stress, *C. zofingiensis* accumulates astaxanthin and canthaxanthin in the cytosolic LBs with concomitant degradation of chloroplast membranes (Rise et al. 1994, Bar et al. 1995). Degradation of thylakoids can be observed within 4 h when *C. zofingiensis* was exposed to high light and nitrogen deprivation, and the grana structure of the thylakoids disappeared with drastic decrease of thylakoid membranes after 24 h under stress. After 3 days, the chloroplast of *C. zofingiensis* lost its typical membrane structure (Bar et al. 1995). A thin lipid-rich layer containing the ketocarotenoids was observed at day 3, possibly function as a sunshade filter to reduce light penetration into the chloroplast and thus prevent formation of excess ROS, a mechanism also found in *H. pluvialis* (Hagen et al. 1994, Bar et al. 1995). Like *H. pluvialis*, a very thick cell wall was observed in *C. zofingiensis* cells under stress, which was resistant to concentrated sulfuric acid treatment. The close proximity of the ketocarot-

enoids layer to the cell wall suggested that the secondary carotenoids may be used as substrates for synthesis of sporopollenin in the cell walls (Bar et al. 1995).

The secondary carotenoids accumulated in *C. zofingiensis* under stress are canthaxanthin and astaxanthin in the forms of free, monoesters and diesters of the pigments (Rise et al. 1994). Canthaxanthin, free astaxanthin and monoesters were detected within 4 h flowing nitrogen deprivation and high light stress, while astaxanthin diesters appeared at 12 h. After 3 days, the main secondary carotenoids accumulated in the cell were canthaxanthin (ca. 25%) and astaxanthin monoesters (ca. 50%) (Bar et al. 1995). Interestingly, *C. zofingiensis* grown heterotrophically can also accumulate significant amounts of astaxanthin (Ip and Chen 2005b). A survey of different carbon sources showed that glucose, mannose, fructose, sucrose, galactose and lactose can be used by *C. zofingiensis*, of which glucose and mannose were the best carbon sources for growth and astaxanthin biosynthesis in the dark. A high astaxanthin yield of 32 mg L⁻¹ was achieved in a feed-batch culture of this organism (Sun et al. 2008).

ASTAXANTHIN BIOSYNTHESIS PATHWAYS

Biosynthesis of astaxanthin in *Haematococcus pluvialis*

Isopentenyl pyrophosphate (IPP) is the precursor for carotenoid synthesis (Lichtenthaler 1999). Two distinct pathways for IPP biosynthesis have been found in higher plants: the mevalonate pathway in the cytosol and the non-mevalonate 1-deoxy-D-xylulose-5-phosphate pathway in the chloroplast (DOXP pathway or MEP pathway) (Lichtenthaler et al. 1997). In unicellular green microalgae *H. pluvialis* and *Chlamydomonas reinhardtii*, IPP is believed to be synthesized solely from the non-mevalonate DOXP pathway (Disch et al. 1998). Subsequently, the isopentenyl pyrophosphate isomerase (IPI) catalyzes the isomerization of IPP to dimethylallyl diphosphate (Lichtenthaler 1999). Introduction of a heterologous *IPI* gene in *Escherichia coli* enhanced the accumulation of carotenoids, supporting a role of IPI in regulating carotenoid biosynthesis (Kajiwarra et al. 1997, Wang et al. 1999). Two cDNAs of IPI, *IPI1* and *IPI2* have been cloned and characterized in *H. pluvialis* (Sun et al. 1998). Transcripts of both *IPI* genes increased in response to oxidative stress, but only *IPI2* was up-regulated at the translational level. Moreover, only *IPI2* protein was detected in mature red cysts in which astaxanthin was accumulated, suggesting

that *IPI2* is responsible for synthesis of the secondary carotenoids, whereas the *IPI1* is responsible for primary carotenoid synthesis in the chloroplast of *H. pluvialis* (Sun et al. 1998).

Phytoene synthase (PSY) catalyzes the first committed step for carotenoid biosynthesis through condensation of two 20-carbon geranylgeranyl pyrophosphate molecules to form a 40-carbon phytoene, the precursor for all other carotenoids (Cunningham and Gantt 1998). Two classes of PSY were found in certain green algae like *Ostreococcus* and *Micromonas*, while some other green algae like *C. reinhardtii* and *C. vulgaris* only possess one class of PSY (Tran et al. 2009a). One copy of *PSY* gene has been cloned and characterized from a number of microorganisms including *H. pluvialis* (Steinbrenner and Linden 2001) and *C. zofingiensis* (Cordero et al. 2011). The *Haematococcus* PSY has an N-terminal extension similar to a chloroplast targeting sequence, indicating that PSY is likely to be targeted to the chloroplast in *H. pluvialis* (Steinbrenner and Linden 2001).

Through a series of dehydrogenation reactions, two structurally similar enzymes, phytoene desaturase (PDS) and ζ -carotene desaturase (ZDS) convert the colorless phytoene into red lycopene. Specifically, PDS catalyzes the first two dehydrogenation reactions to form phytofluene and ζ -carotene, whereas ZDS catalyzes two further reactions converting ζ -carotene to neurosporene and lycopene (Cunningham and Gantt 1998). These dehydrogenation reactions extend the conjugated carbon-carbon double bonds to form the chromophore of carotenoids. These FAD-containing enzymes require PTOX and plastoquinone (PQ) as electron acceptors (Carol et al. 1999, Wu et al. 1999). The *PDS* and two *PTOX* genes (i.e., *PTOX1* and *PTOX2*) have been cloned and characterized in *H. pluvialis* (Grunewald et al. 2000, Wang et al. 2009, Li et al. 2010). High light illumination and nitrogen deprivation increase the transcripts of *PDS* and *PTOX* simultaneously in *H. pluvialis*, suggesting that PDS and PTOX may act in concert to dehydrogenate phytoene and remove excess electrons under stress, thereby preventing over-reduction of the photosynthetic electron transport chain and the formation of excess ROS (Grunewald et al. 2000, Wang et al. 2009, Li et al. 2010). Western-blot and immunogold labeling experiments showed that PDS is located exclusively in the chloroplast in *H. pluvialis* (Grunewald et al. 2000).

The cyclization of lycopene catalyzed by lycopene β -cyclase (LCY-b) and lycopene ϵ -cyclase (LCY-e) is a branching point in the carotenoid biosynthesis (Cunningham and Gantt 1998). LCY-b catalyzes two β -cyclization

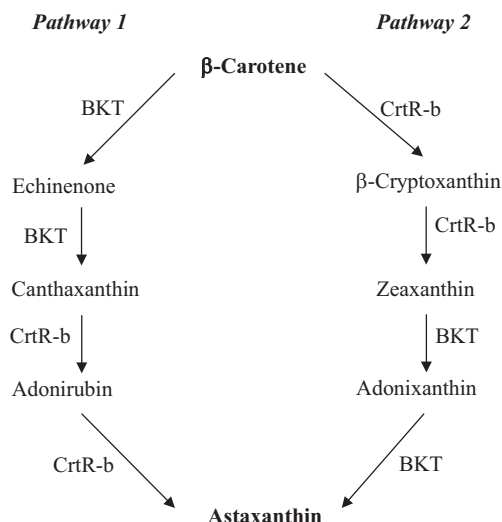


Fig. 1. Primary astaxanthin biosynthesis pathways in *Haematococcus pluvialis* (pathway 1) and *Chlorella zofingiensis* (pathway 2). Enzymes are named according to the designation of their genes. BKT, β -carotenoid ketolase; CrtR-b, β -carotenoid hydroxylase. Modified and reproduced from Li (2007) with permission from the University of Hong Kong.

reactions at each end of lycopene to form β -carotene, which is an end product for photosynthesis in plants and algae, as well as a precursor for keto-carotenoids in the chloroplast and cytosol. LCY-e catalyzes ϵ -cyclization of lycopene to ζ -carotene, which is subsequently cyclized by LCY-b to form α -carotene (β,ϵ -carotene), a precursor for lutein synthesis (Cunningham et al. 1996). A *LCY-b* gene has been cloned in *H. pluvialis* and was observed that its transcripts increased concomitantly with the formation of β -carotene and astaxanthin at increased intensities of red or blue light (Steinbrenner and Linden 2003).

Oxygenation of β -carotene at the 4 position by β -carotene ketolase (BKT, synonym *CrtW*) gives rise to echinenone and canthaxanthin (Fig. 1). In *H. pluvialis*, three different *BKT* genes have been cloned, including *BKT1* (Lotan and Hirschberg 1995), *BKT2* (Kajiwar et al. 1995), and *BKT3* (Huang et al. 2006a). Under stress, the multiple *BKT* genes were up-regulated and when total *BKT* mRNAs reached a certain threshold, *H. pluvialis* began to synthesize astaxanthin (Huang et al. 2006a). Immuno-localization study indicated that BKTs were localized in both the chloroplast and cytosolic LBs in *H. pluvialis*, although ketolase activity was detected only in the cytosolic LB preparation (Grünwald et al. 2001). Since PDS is an early enzyme in the pathway that is located exclusively in the chloroplast (Grünwald et al. 2000), it was proposed that transport of carotenoid precursors from the chloroplast to cytosolic LBs by a yet unknown

mechanism would be necessary for astaxanthin formation (Grünwald et al. 2001).

Heterologously expressed *Haematococcus BKT1* was not able to use dihydroxy carotenoid zeaxanthin as a substrate, indicating that the oxygenation steps likely precede hydroxylation steps (Lotan and Hirschberg 1995). *In vitro* assay of BKT and β -carotene hydroxylase (CrtR-b, synonym Chy-b) further confirmed that astaxanthin synthesis in *H. pluvialis* is preferably preceded with the addition of keto groups, followed by the hydroxylation reactions (Fraser et al. 1998, Schoefs et al. 2001). Astaxanthin in *H. pluvialis* is proposed to be synthesized from the hydroxylation of canthaxanthin catalyzed by CrtR-b (Fig. 1). *CrtR-b* gene has been isolated in *H. pluvialis* (Linden 1999). A linear relationship between the increase of *CrtR-b* transcripts and astaxanthin concentration was observed in the *H. pluvialis* wild type and the astaxanthin-hyper-accumulating mutant (MT 2877), suggesting a transcriptional control of *CrtR-b* over astaxanthin synthesis (Li et al. 2008b). BKT and CrtR-b have been often proposed as rate-limiting steps in astaxanthin synthesis. Genetic manipulation of their genes in algae and plants for enhanced astaxanthin production is further discussed in the BIOTECHNOLOGICAL IMPLICATIONS section.

Regulation of carotenogenesis in *Haematococcus pluvialis*

Most *Haematococcus* carotenoid synthesis genes are up-regulated at the transcriptional level in response to various stressors (e.g., high light, excess ferrous sulfate and excess sodium acetate) (Li et al. 2008b). The extent to which the individual stressor affecting carotenogenesis gene expression follows the order as: low light < low light + ferrous sulfate < low light + sodium acetate < high light + ferrous sulfate/sodium acetate < high light + ferrous sulfate + sodium acetate (Li et al. 2008b). Interestingly, when multiple factors were applied simultaneously to *H. pluvialis*, for example, high light + salt stress + iron stress, the transcripts of carotenoid synthesis genes were lower than and the maximum expression levels of those genes were delayed compared to that induced by the individual stressors applied separately or two stressors together at the early stage of stress (Li et al. 2008b). A plausible interpretation was that *H. pluvialis* and other microalgae possess both “house-keeping” defense mechanisms and stressor-specific defense mechanisms. For example, the carotenogenesis for production of astaxanthin and other secondary carotenoids is largely induced under high light stress, whereas various enzymatic defense reactions may

be triggered by salinity or excess iron ion. When multiple stressors were applied to *H. pluvialis*, multiple stress protection mechanisms may all be activated and work in concert, each contributing partially to overall cell protection and to a less extent carotenogenesis. However, because the enzymatic defense systems mainly serve as short-term cellular defense mechanisms in *H. pluvialis* (Wang et al. 2004a), as the stress persists, the cells would have to depend more upon the long-term defense mechanism, β -carotenogenesis, for survival, which explained why a delayed expression pattern of carotenogenesis genes with greater expression levels occurred in the cultures treated with the multiple stressors (Li et al. 2008b).

ROS play a role in regulation of carotenogenesis in *H. pluvialis*. Treatment with ROS-generating compounds such as methylviologen, methylene blue and Fe^{2+} , increased *PDS* and *CrtR-b* expression in *H. pluvialis*. They also induced astaxanthin synthesis independent of *de novo* protein synthesis (Kobayashi et al. 1993, Steinbrenner and Linden 2001), suggesting ROS mediate stress response and carotenogenesis in *H. pluvialis* at a post-translational level (Steinbrenner and Linden 2003). Unlike *PDS* and *CrtR-b*, however, treatment with various transcriptional and translational inhibitors suggested that *BKT* gene expression was dependent on *de novo* protein synthesis (Vidhyavathi et al. 2008). Considering *PDS* is shown to be located in the chloroplast, whereas *BKT* activity is found in the cytosolic LBs, this discrepancy can be explained by the distinct subcellular localization of individual carotenoid biosynthetic enzymes.

Pathways and regulation of carotenogenesis in *Chlorella zofingiensis*

C. zofingiensis has the identical β -carotene synthesis pathway found in *H. pluvialis*, but may take a different route to make astaxanthin (Fig. 1) (Li 2007). Recently, the genes involved in biosynthesis of astaxanthin in *C. zofingiensis* have been cloned and characterized, including *PSY* (Cordero et al. 2011), *PDS* (Huang et al. 2008), *LCY-b* (Cordero et al. 2010), *LCY-e* (Cordero et al. 2012), *BKT* (*CrtO*) (Huang et al. 2006b), *CrtR-b* (*Chy-b*) (Li et al. 2008a). Under high-light conditions, *PSY*, *PDS*, *BKT*, *CrtR-b* genes were up-regulated, whereas the mRNA levels of *LCY-b* and *LCY-e* remained constant, leading to formation of secondary carotenoids (Li et al. 2009, Cordero et al. 2012).

Functional analysis of *C. zofingiensis* *BKT* demonstrated that this enzyme not only converted β -carotene to canthaxanthin via echinenone, but also exhibited high

enzymatic activity of converting zeaxanthin to astaxanthin via adonixanthin (Huang et al. 2006b). When the *BKT* inhibitor diphenylamine was applied, *C. zofingiensis* accumulated zeaxanthin while *H. pluvialis* accumulated β -carotene (Wang and Chen 2008), suggesting that *C. zofingiensis* synthesizes astaxanthin through zeaxanthin (Fig. 1, Pathway 2).

Various environmental factors may differentially regulate carotenogenesis in *C. zofingiensis*. High irradiance triggered up-regulation of *PDS*, *BKT*, *CrtR-b* genes in *C. zofingiensis*, with most significant increase in *CrtR-b* transcripts, leading to the accumulation of canthaxanthin, zeaxanthin and astaxanthin (Li et al. 2009). Under salt stress, only *BKT* gene was up-regulated and canthaxanthin and astaxanthin were accumulated (Li et al. 2009). When fed with sugars, *C. zofingiensis* accumulated astaxanthin in dividing cells. Glucose and mannose were the best carbon sources to sustain growth and astaxanthin production in *C. zofingiensis*, followed by fructose, sucrose and galactose, whereas lactose was the poorest carbon source for *C. zofingiensis* (Sun et al. 2008). In heterotrophically grown *C. zofingiensis*, carotenogenesis was regulated through the phosphorylation of glucose by hexokinase, which was essential for up-regulation of *CrtR-b* and synthesis of zeaxanthin, whereas the signals from the mitochondrial alternative pathway may mediate up-regulation of *BKT* and synthesis of astaxanthin (Li et al. 2008a).

Two possible types of signals from the mitochondrial alternative pathway may regulate carotenogenesis in *C. zofingiensis*: ROS and organic acids from the tricarboxylic acid cycle (TCA) cycle. Chemically generated ROS induce carotenogenesis and astaxanthin formation in *C. zofingiensis* (Ip and Chen 2005a, Li et al. 2009). Singlet oxygen specifically induced *CrtR-b* whereas hydroxyl radical up-regulated *PDS* and *BKT*, suggesting their distinct roles in regulating carotenogenesis in *C. zofingiensis* (Ip and Chen 2005a). In corroboration, the fatty acid synthesis genes *SAD* (coding for stearyl acyl carrier protein) and *BC* (coding for biotin carboxylase) seemed to be associated with ROS (Liu et al. 2012). On the other hand, TCA cycle acids such as citrate may act as a signal at the gene expression level to induce mitochondrial alternative pathway respiration to facilitate carotenogenesis (Vanlerberghe and McIntosh 1996). In *C. zofingiensis*, citrate was shown to induce *BKT* expression and astaxanthin synthesis independent of ROS formation, suggesting that the mitochondrial alternative pathway mediates *BKT* expression through modulation of TCA cycle with citrate as a signal molecule (Li et al. 2008a).

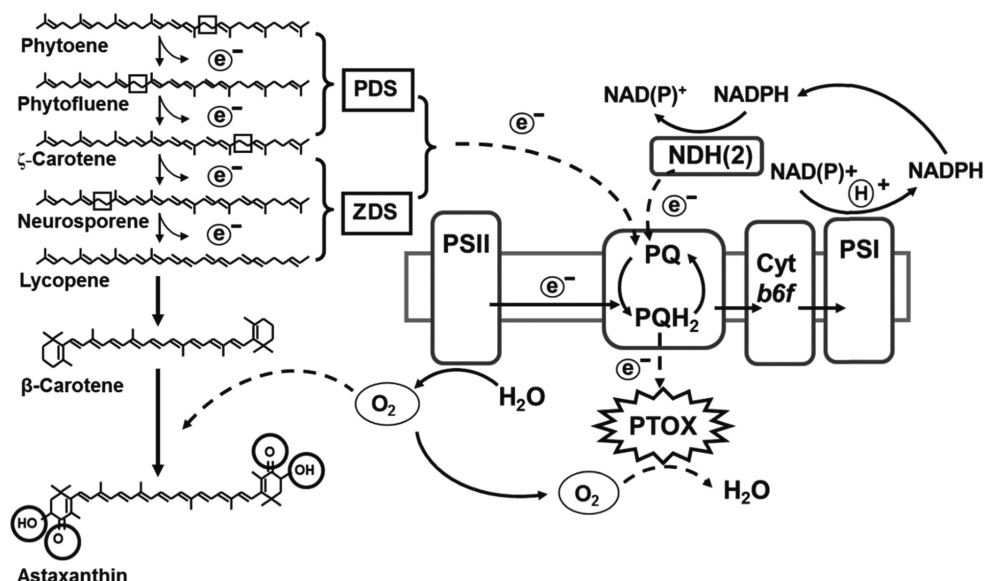


Fig. 2. Multiple roles of the astaxanthin biosynthesis in protecting microalgae against oxidative stress. Astaxanthin biosynthesis pathway functions to 1) reduce subcellular oxygen levels via synthesis of oxygen-rich astaxanthin and its esters; 2) convert photosynthetically evolved oxygen into water via a coupled electron transport from carotenoid desaturation steps (phytoene desaturase [PDS] and ζ-carotene desaturase [ZDS]) to the plastoquinone pool to plastid terminal oxidase (PTOX), whereas astaxanthin itself; 3) serves as a “sunshade” to reduce excess light illumination on the photosystems; 4) functions as a powerful antioxidant against reactive oxygen species. PS, photosystem; PQ, plastoquinone. Modified and reproduced from Li (2007) with permission from the University of Hong Kong.

Crosstalk between astaxanthin synthesis pathway and photosynthetic electron transfer chain

In *H. pluvialis*, the nuclear carotenoid biosynthetic genes were suggested to be regulated by the redox state of the PQ pool at the photosynthetic electron transfer chain, as carotenoid biosynthesis genes are up-regulated under high PSII excitation pressure (PQ reduced) (Steinbrenner and Linden 2003), and the inhibition of photosynthesis by photosynthetic electron transport inhibitor DCMU abolished the up-regulation of the *PSY* gene induced by high light (Steinbrenner and Linden 2001). However, how the signal of redox state of the photosynthetic electron transport chain can pass the chloroplast envelope and regulate nuclear gene expression is not known. It was proposed that protein phosphorylation cascade may be a possible mechanism to mediate this retrograde signaling process (Chandok et al. 2001, Chen et al. 2004). Furthermore, the chlorophyll biosynthesis precursors, Mg-protoporphyrin IX and Mg-protoporphyrin-methylesters, were proposed to play a role in chloroplast-to-nucleus retrograde signaling (Nott et al. 2006).

Besides the redox state of PQ, other photosynthetic signals such as ROS, trans-thylakoid pH gradient and the stoichiometry of PSI and PSII may also contribute to the photosynthetic regulation of carotenogenesis gene ex-

pression (Fey et al. 2005). Recently, a transcriptomic analysis has identified over 2,000 genes regulated under high irradiance conditions, providing therefore more candidate genes for further study of the crosstalk between photosynthesis and carotenogenesis gene expression in *H. pluvialis* (Kim et al. 2011).

In addition to the regulation at the gene expression level, metabolic coupling between carotenoid and photosynthetic electron transport may be present in *H. pluvialis*. The electrons produced in the sequential desaturation reactions catalyzed by PDS and ZDS can be delivered to the PQ pool, which is in turn oxidized by PTOX (Fig. 2). Two genes encoding PTOX (*PTOX1* and *PTOX2*) in *H. pluvialis* have been cloned and *PTOX1* was found to be co-regulated with the carotenoid biosynthesis genes under various environmental stress conditions (Li et al. 2008b, 2010, Wang et al. 2009). Functional analysis of PTOX of *Chlamydomonas* also suggested that PTOX1 was responsible for regenerating oxidized PQ for phytoene desaturation (Houille-Vernes et al. 2011).

Crosstalk between astaxanthin and fatty acid biosynthesis pathways

In *H. pluvialis*, over 90% astaxanthin molecules are esterified with fatty acids (Yuan et al. 1997). Oleic acid is

the major fatty acid species that is conjugated to astaxanthin molecules (Holtin et al. 2009). A linear correlation between the cellular fatty acid content and astaxanthin content in *H. pluvialis* under stress conditions has been reported in several independent studies (Zhekisheva et al. 2002, Chen 2007), leading to a hypothesis that the biosynthesis of astaxanthin and fatty acids are coupled in this organism. This hypothesis was further supported by the fact that with the addition of the fatty acid synthesis inhibitor cerulenin or the carotenoid synthesis inhibitor norflurazon, both astaxanthin and fatty acid biosynthesis were abolished (Schoefs et al. 2001, Chen 2007). Astaxanthin esterification must be the reaction that links the two pathways. Acyl-CoA, the presumable substrate for astaxanthin esterification, when in excess, may feedback-inhibit acetyl-CoA carboxylase, a rate-limiting enzyme for fatty acid synthesis (Ohlrogge and Jaworski 1997). Presumably, the inhibition of astaxanthin synthesis by norflurazon may result in building up relatively excess amounts of acyl-CoA which may in turn inhibit fatty acid synthesis through a feedback inhibition mechanism. Likewise, the enzymes responsible for astaxanthin synthesis may also be under feedback control when fatty acid biosynthesis is inhibited by cerulenin. Astaxanthin esterification, the key point for flux control, needs to be further characterized at the gene, enzyme and subcellular levels.

Physiological role of astaxanthin biosynthesis

In algae and higher plants, ROS are mainly generated through photosynthesis in chloroplasts (Asada 2006). Under stress conditions, e.g., high light, nutrient starvation, and high salinity, the imbalance between generation and detoxification of ROS within the chloroplast may cause photooxidative stress.

Astaxanthin was proposed to act as a “sunshade” to reduce the penetration of blue light into the chloroplast, thereby reducing photooxidative damage of the photosystems by excessive light (Hagen et al. 1994). Astaxanthin could also act as a physical-chemical barrier to prevent DNA, RNA, enzymes and membrane lipids from ROS attack (Hagen et al. 1993, Kobayashi et al. 1997, Kobayashi and Sakamoto 1999). Astaxanthin esters were shown to scavenge ROS, e.g., the superoxide anion radicals (O_2^-) and singlet oxygen (1O_2) and thus may act as an antioxidant agent against ROS (Kobayashi et al. 1997, Kobayashi and Sakamoto 1999).

The photoprotective role of astaxanthin has been challenged since astaxanthin-rich cysts of *H. pluvialis* were

equally or more susceptible to high light stress than astaxanthin-free vegetative cells (Fan et al. 1998). These authors suggest that the multiple intermediate reactions in astaxanthin synthesis pathway may consume excessive electrons and thus prevent over-reduction of the photosynthetic electron transport chain and reduce the production of ROS. As such, astaxanthin was proposed as an end-product rather than the protective agent itself in the stress response process (Fan et al. 1998).

More recently, Li et al. (2008b) proposed that astaxanthin biosynthesis exerts multilevel protective roles against photooxidative stress. In addition to the protective roles summarized above, astaxanthin biosynthesis protects *H. pluvialis* cells from stress through the consumption of molecular oxygen (Li et al. 2008b). Up to 10% of molecular oxygen evolved from photosynthesis under stress is consumed by astaxanthin synthesis via two distinct routes: 1) use oxygen as a substrate for ketocarotenoids formation and 2) convert oxygen generated from photosynthesis and electrons from the carotenoid desaturation steps to water by PTOX connected to the PQ pool. Decrease in subcellular molecular oxygen concentration reduces the substrate available for oxygen dependent ROS formation, as is the case for the alternative oxidase which reduces the O_2 concentration and prevents electrons from reducing O_2 to $O_2^{\cdot-}$ (Mittler 2002). Re-oxidation of the PQ pool by PTOX may further relax the photosynthetic electron transport chain and reduce ROS formation (Li et al. 2008b). The multiple roles of the astaxanthin biosynthesis in protecting microalgae against oxidative stress are summarized in Fig. 2 (Li 2007).

In addition to the ability to accumulate large amounts of astaxanthin, microalgae such as *H. pluvialis* have evolved other mechanisms for ROS scavenging, for example, by the antioxidative enzyme defense system. Proteomics analysis revealed several superoxide dismutases (SOD), peroxidases, and catalases were up-regulated in response to photooxidative stress (Wang et al. 2004a, Tran et al. 2009b). Transcriptomic analysis revealed that the up-regulation of SOD, catalase, and glutathione peroxidase genes occurred under high light and nutrient starvation conditions (Eom et al. 2006, Kim et al. 2011, Wang et al. 2011). All the antioxidative enzymes studied showed a transient up-regulation pattern and then reverted to the basal or below basal level, suggesting that the enzyme defense system is an early response mechanism to oxidative stress, and as stress persists, the cells will adopt astaxanthin biosynthesis and accumulation as a long-term defense mechanism (Wang et al. 2004a).

BIOTECHNOLOGICAL IMPLICATIONS

Metabolic engineering for enhanced carotenoid production

Many higher plants exhibit enzymatic activity of carotenoid hydroxylase but not β -carotene ketolase, and as a result, they produce 3-hydroxy carotenoids (e.g., lutein and zeaxanthin) but no 4-ketocarotenoids such as astaxanthin. Introduction of a *Haematococcus* β -carotene ketolase gene (*BKT*) into a tobacco plant resulted in moderate accumulation of astaxanthin (ca. 840 $\mu\text{g g}^{-1}$ dwt) in the nectary tissue (Mann et al. 2000). A similar effort was made with a carrot plant, resulting in a comparable amount (ca. 916 $\mu\text{g g}^{-1}$ dwt) of astaxanthin in root tissues (Jayaraj et al. 2008). Expression of a *Chlamydomonas* *BKT* into *Arabidopsis* led to the accumulation of greater amounts of astaxanthin (up to 2,000 $\mu\text{g g}^{-1}$ dwt) in leaves of the transgenic plant, whereas the expression of *C. zoofingensis* *BKT* only resulted in 240 μg astaxanthin g^{-1} dwt (Zhong et al. 2011). A greater amount of astaxanthin (1,600 $\mu\text{g g}^{-1}$ dwt) in leaves of a transgenic tobacco was also obtained when the *Chlamydomonas BKT* was introduced (Huang et al. 2012), suggesting that the catalytic capacity of *BKT* is species-specific (Zhong et al. 2011). A much greater amount of astaxanthin (5,440 $\mu\text{g g}^{-1}$ dwt) was obtained in tobacco leaves when a *BKT* and a *CrtR-b* gene from a marine bacterium *Brevundimonas* sp. were co-transformed into the plant plastid genome (Hasunuma et al. 2008).

Lack of dominant selectable markers has hindered the genetic engineering of astaxanthin production in microalgae. Recently, a stable nuclear transformation system has been established in *H. pluvialis* (Steinbrenner and Sandmann 2006) and in *C. zoofingensis* (Huang et al. 2008). A modified version of *PDS* gene in *H. pluvialis* and *C. zoofingensis* has been developed as a dominant selectable marker and was transformed into these two species through biolistic transformation (Steinbrenner and Sandmann 2006, Huang et al. 2008, Liu et al. 2009). *Agrobacterium*-mediated transformation has also been reported to express foreign genes in *Haematococcus* (Kathiresan et al. 2009). The newly developed genetic transformation tools for *H. pluvialis* and *C. zoofingensis* will provide metabolic engineering opportunities for overproduction of astaxanthin in these two and perhaps other microalgae. One possible application is to enhance astaxanthin production in *H. pluvialis* or *C. zoofingensis* by overexpressing *PSY* and *CrtR-b* genes, which previously have been shown to be possible rate-limiting steps for astaxanthin synthesis in *H. pluvialis* (Li et al. 2008b, 2010).

An alternative strategy for enhancing astaxanthin production in microalgae is through random mutagenesis. Through chemical mutagenesis, several *Haematococcus* mutants with improved phenotypic traits have been generated, and one such example is *Haematococcus* astaxanthin-hyper-accumulating mutant strain MT 2877 (Hu et al. 2008). In the early vegetative growth stage, MT 2877 was identical to the wild type with respect to cell morphology and physiology. However, the mutant grew faster under stress conditions with about 100% more viable cells. MT 2877 also accumulated 100% more astaxanthin, leading to a 4-fold increase in volumetric astaxanthin productivity compared to the wild type (Hu et al. 2008). Earlier, two *H. pluvialis* cell wall deficient mutants, MT 537 and MT 2978, have been generated by chemical mutagenesis (Wang et al. 2005). *Haematococcus* wild type cells possess thick and rigid cell walls that reduce considerably the bioavailability of astaxanthin to human and animals if whole *Haematococcus* cells are consumed without cell disruption by chemical or physical means. The cell wall mutants were demonstrated to significantly improve the bioavailability of astaxanthin. Production of astaxanthin using the cell wall mutants may also reduce downstream processing costs whereby eliminating or minimizing the cell wall disruption step which is otherwise required for wild type *Haematococcus* (Wang et al. 2005, Hu et al. 2006). Some mating-based breeding techniques can also be applied to *Haematococcus* to improve existing or introduce new desirable traits, given that many *Haematococcus* strains vary in photosynthesis efficiency, growth, astaxanthin content, and susceptibility to oxidative stress or parasite attack. Any efforts along this line would be valuable.

Effects of environmental factors on growth and astaxanthin production

Growth and carotenogenesis of *H. pluvialis* are regulated by various environmental factors, such as light, temperature and nutrients. The maximal specific growth rate of *H. pluvialis* is 0.054 h^{-1} , corresponding to a doubling time of 12–13 h. Such a high growth potential occurs only under favorable growth conditions, e.g., low light irradiance (20–50 $\mu\text{E m}^{-2} \text{s}^{-1}$), optimal temperature (25–28°C) and replete nutrients (Fan et al. 1994). On the contrary, high light, high temperature, and nutrient deprivation (e.g., nitrogen and phosphorus) induce astaxanthin synthesis while retarding cell division. High light is one of the most effective factors to stimulate astaxanthin synthesis and thus is often applied to increase astaxanthin produc-

tion (Choi et al. 2002). High temperature is rarely implemented to induce astaxanthin production, as it was reported to severely reduce biomass yield and thus overall reduced astaxanthin productivity (Tjahjono et al. 1994).

Carotenogenesis in *H. pluvialis* can be stimulated by a variety of metal ions such as Fe^{2+} , Mn^{2+} , and Cd^{2+} (Harker et al. 1996b). All the tested metals exerted inhibitory effect on cell growth except Fe^{2+} . Fe^{2+} was suggested to stimulate ROS (especially hydroxyl radicals, $\text{HO}\cdot$) production via the Fenton reaction, which in turn trigger astaxanthin synthesis (Tjahjono et al. 1994). This speculation was supported by several lines of evidence: EDTA-chelated $\text{FeCl}_3\cdot 6\text{H}_2\text{O}$ (a form of iron which does not cause Fenton reaction) did not induce astaxanthin synthesis, whereas potassium iodide with a capability of scavenging $\text{HO}\cdot$ and thus abolishing Fe^{2+} -induced astaxanthin synthesis in *H. pluvialis* (Boussiba and Vonshak 1991, Kobayashi et al. 1993).

Astaxanthin synthesis in *H. pluvialis* can also be induced by salinity stress. NaCl (0.1-0.5%, w/w) was used to increase astaxanthin accumulation in laboratory cultures (Harker et al. 1996a, Sarada et al. 2002). However, relatively high concentrations of NaCl (0.6-0.8%, w/w) may cause severe cell mortality, in particularly for flagellated zoospores, which limits the implementation of this strategy in large-scale *Haematococcus* culture (Harker et al. 1996a, Sarada et al. 2002, Cifuentes et al. 2003).

Mass cultivation of *Haematococcus pluvialis* and *Chlorella zofingiensis*

Photoautotrophic culture. Photoautotrophic culture of *H. pluvialis* is carried out in open raceway ponds and closed photobioreactors (Olaizola 2000, Carvalho et al. 2006, Ugwu et al. 2008). Because the culture conditions for maximum growth and maximum astaxanthin content are mutually exclusive, a two-stage batch culture mode is commonly adopted for mass cultivation of *H. pluvialis*. In the first stage (also called 'green stage' because the cells are green), the cells are maintained in a nutrient-replete growth medium and exposed to low light intensity to promote biomass production. When the cells enter into the stationary growth phase, the culture is then transited into the second stage (also called 'red stage' because the cells are turning red from green) where the cells are subjected to high light intensity and nutrient deprivation to induce astaxanthin production. The reported biomass productivities range from 0.03 to 0.6 $\text{g L}^{-1} \text{d}^{-1}$ at the green stage and from 0.01 to 0.58 $\text{g L}^{-1} \text{d}^{-1}$ at the red stage, and astaxanthin productivity ranged from 0.4 to 17 $\text{mg L}^{-1} \text{d}^{-1}$

(Boussiba et al. 1992, Olaizola 2000, Fábregas et al. 2001, Aflalo et al. 2007).

During the first two days of the red-stage, considerable cell death (or photo-bleach) occurs, primarily to flagellated zoospores, due to their higher susceptibility to photooxidative stress than non-motile palmella cells (Harker et al. 1996a, Sarada et al. 2002, Han et al. 2012). To prevent or reduce cell mortality caused by high light and nutrient deprivation, the initial cell concentration in the culture must be optimized (Wang et al. 2013). The highest astaxanthin productivity of 17.5 $\text{mg L}^{-1} \text{d}^{-1}$ was obtained at the red stage where an optimal initial biomass density of 0.8 $\text{g L}^{-1} \text{dwt}$ was applied to an outdoor glass column photobioreactor (Wang et al. 2013). Greater tolerance of palmella than zoospores to photooxidative stress resulted in more than 10-fold increase in astaxanthin productivity (Choi et al. 2011).

Besides the two-stage culture mode, a single-stage cultivation mode has also been tested to produce astaxanthin in flagellates of some *H. pluvialis* strains in a chemostat system (Del Río et al. 2005, 2008, García-Malea et al. 2009). Under optimal light irradiance, nutrient concentration and dilution rate, algal biomass productivities of 0.7-1.9 $\text{g L}^{-1} \text{d}^{-1}$ were obtained, corresponding to an astaxanthin productivity of 5.6-21 $\text{mg L}^{-1} \text{d}^{-1}$. The technical and economic feasibilities of this single-stage culture mode for mass culture of *H. pluvialis* remain to be seen.

Although a number of *Chlorella* species and strains have been tested for carotenoid production on laboratory scales, little has been done on a large-scale (Liu et al. 2011).

Heterotrophic and mixotrophic culture modes. *H. pluvialis* is capable of utilize organic carbon for growth in the absence of light, which provides the means of heterotrophic and mixotrophic cultivation for astaxanthin production. Under heterotrophic conditions, *H. pluvialis* grows at a relatively low growth rate (0.22 d^{-1}) and accumulates ca. 0.5% dwt of astaxanthin (Kobayashi et al. 1992). Growth and astaxanthin production can be enhanced under mixotrophic culture conditions. A final cell density of 0.9-2.65 g L^{-1} and a maximum astaxanthin content of 1-2% dwt were obtained from mixotrophic cultures of *H. pluvialis* (Chen et al. 1997, Zhang et al. 1999, Wang et al. 2004a). A heterotrophic-photoautotrophic culture mode was also explored where heterotrophic culture produced algal biomass, while astaxanthin production was induced during photoautotrophic culture. As a result, a very high cellular astaxanthin content of 7% by dwt, but low astaxanthin productivities of 4.4-6.5 $\text{mg L}^{-1} \text{d}^{-1}$ were obtained (Hata et al. 2001, Kang et al. 2005).

Much higher cell density can be achieved in heterotrophic culture of *C. zofingiensis*. For example, high cell concentrations of 10 g L⁻¹ (Ip and Chen 2005b) and 53 g L⁻¹ (Sun et al. 2008) were achieved in *C. zofingiensis* cultures fed with glucose with a maximum specific growth rate of 1.03 d⁻¹. A moderate astaxanthin yield of 12.5 mg L⁻¹ was obtained in a mixotrophic culture of *C. zofingiensis* (Ip et al. 2004), and the yield was increased to 32 mg L⁻¹ in a glucose fed-batch culture. However, the astaxanthin productivity is low ranging from 0.9 to 2 mg L⁻¹ d⁻¹ (Sun et al. 2008).

Microbial contamination and crop protection. Mass culture of *Haematococcus* can be contaminated by fungal parasites and zooplanktonic predators (e.g., amoebas, ciliates, and rotifers), as well as other microalgae and cyanobacteria, resulting in reduced biomass yield and quality, and sometimes loss of culture all together. A parasitic blastoclad fungus identified as *Paraphysoderma sedebokerensis* is the most devastating disease responsible for reduced astaxanthin productivity and frequent culture collapses (Hoffman et al. 2008, Gutman et al. 2009). Hoffman et al. (2008) made detailed observations on the infection of *Haematococcus* culture by the parasite *P. sedebokerensis* (Hoffman et al. 2008). During the infection process, healthy green culture turns dark brown, accompanied by the formation of large clumps consisting of living cells, cell debris, and particulate organic matters. A more vivid red color of culture can be first observed if infection occurred at the red stage, followed by gradual bleaching of algal cells (Hoffman et al. 2008).

The infection process can be somewhat inhibited in the presence of lectin-like carbohydrate-binding proteins, which were assumed to bind to the surface of palamelloids and aplanospores of *Haematococcus* (Gutman et al. 2011). The recognition between the blastoclad parasite and *Haematococcus* cells was suggested to be a lectin-N-acetyl-D-glucosamine interaction. More research along this line may better understand the molecular and cellular mechanisms by which *P. sedebokerensis* recognizes and infects *Haematococcus*, which may in turn lead to develop a biology-inspired method to prevent or treat the fungal disease in the mass culture of *Haematococcus* or other related species.

Downstream processes and product formation

Haematococcus aplanospores can be readily harvested by sedimentation and centrifugation prior to spray. If whole *Haematococcus* cells are a final form of product, *Haematococcus* pastes from centrifugation is usually

pretreated by a mechanical means to disrupt the rigid cell walls, followed by a spray-drying process to produce dry biomass powder. If astaxanthin is the final product, a conventional solvent-based extraction method or supercritical fluid extraction can be applied to wet or dry *Haematococcus* biomass to obtain concentrated astaxanthin extracts (Bubrick 1991, Mendes-Pinto et al. 2001, Nobre et al. 2006, Sarada et al. 2006, Krichnavaruk et al. 2008).

CONCLUSIONS AND PERSPECTIVES

The occurrence and accumulation of astaxanthin in microalgae is a strategy to cope with oxidative stress. Astaxanthin synthesis pathways interact with multiple metabolic pathways such as the photosynthetic electron transport, fatty acid synthesis, and ROS generation. However, the molecular mechanisms mediating the crosstalk between astaxanthin synthesis and related metabolic pathways remain elusive. Recent progress in genome sequencing and genetic toolbox development of microalgae has made it possible to further address these questions. The new knowledge obtained from the studies of astaxanthin biosynthesis and stress response will be applied for enhanced astaxanthin production. Until now, *H. pluvialis* is still the best producer for astaxanthin although the biomass productivity is relatively low. Future efforts should focus on increasing biomass production of *H. pluvialis* or screening for and manipulating other fast-growing algal strains for enhanced astaxanthin production along with improved phenotypic traits for various commercial applications.

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